

(19)日本国特許庁 (J P)

(12) 公表特許公報 (A)

(11)特許出願公表番号

特表平11-501504

(43)公表日 平成11年(1999)2月9日

(51)Int.Cl.⁸
C 1 2 N 15/09

識別記号
Z N A

F I
C 1 2 N 15/00

Z N A A

審査請求 未請求 予備審査請求 未請求(全 50 頁)

(21)出願番号 特願平8-518463
(86) (22)出願日 平成7年(1995)12月12日
(85)翻訳文提出日 平成9年(1997)6月12日
(86)国際出願番号 P C T / G B 9 5 / 0 2 8 9 3
(87)国際公開番号 W O 9 6 / 1 8 7 3 1
(87)国際公開日 平成8年(1996)6月20日
(31)優先権主張番号 9 4 2 5 1 3 8 . 6
(32)優先日 1994年12月12日
(33)優先権主張国 イギリス (G B)

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スタッドファイエン 93
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最終頁に続く

(54)【発明の名称】 核酸の単離方法

(57)【要約】

本発明は、サンプルを界面活性剤および固体支持体に接触させ、これによって上記サンプル中の可溶性核酸がこの支持体に結合し、次いでこの支持体を結合した核酸とともに上記サンプルから分離することからなる、サンプルから核酸を単離する方法を提供する。本発明の方法を、DNA を単離するのに用いる場合、同一のサンプルから RNA を単離する追加工程と都合良く組み合わせることができる。

(2)

RNA

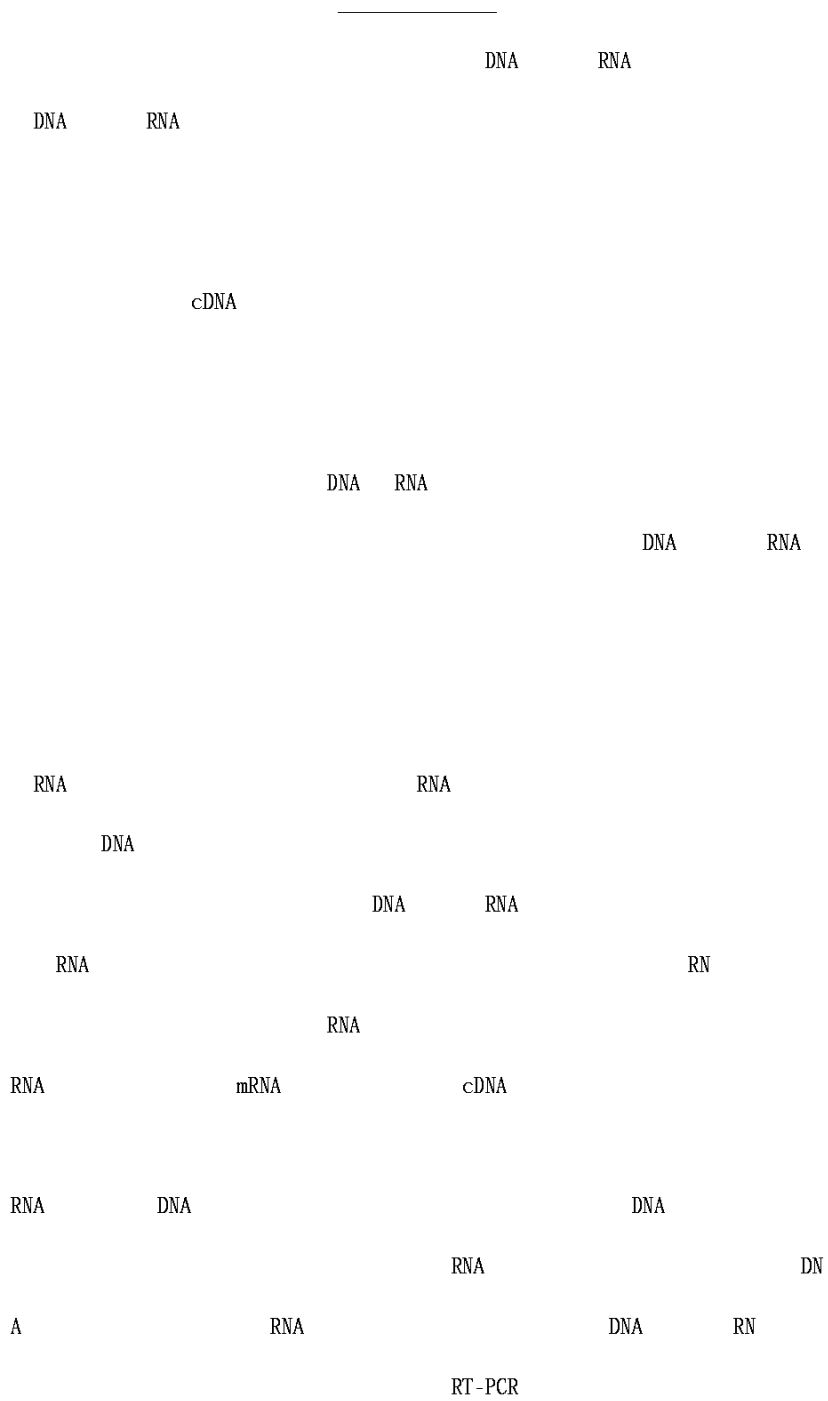
DNA

RNA

0.2 30% w/v

RNA

(4)



NA LiCl DNA R

Cellular Component	DNA	RNA	mRNA
Nucleus	Yes	Yes	No
Mitochondrion	Yes	Yes	No
Cytoplasm	No	Yes	Yes
Ribosome	No	Yes	Yes

	mRNA	50	300	A
	mRNA			(polyadenylated)
(A) + mRNA	RNA	95	%	R
NA	RNA			A

A

RNA

(dT) -

mRNA

(6)

dT

mRNA

(A)⁺ RNA

mRNA

RNA

(A)⁺ RNA

microfuge

RNA

LiCl

LiDS/SDS

dT

-K

mRNA

15

mRNA

30

mRNA

mRNA

mRNA

mRNA

GTC

sarkosyl

GTC-

RN

4M GTC

DNA

US-A-5,234,8

(7)

WO 91/12079

RNA

PCR

PCR

RNA

DNA

DNA RNA

DNA

DNA

cDNA

DNA

RNA

RNA

PCR

(Banerjee S K et al. 1995 Biotechniques 18:769-773)

20

0.5

(SDS)

0.2 30 %

0.5 30 %

0.5 15 %

10 %

1.0 %

0.5 %

			0.1
M	250	500 mM	
			50 mM
	mEDTA	EGTA	
10 mM		DTT	-
100mM	-HCl	pH 7.5	
10mM	EDTA		
2%	SDS		
100mM	C1	pH 7.5	
10mM	EDTA		
5%	SDS		
10mM	NaCl		
100mM	C1	pH 7.5	
500mM	LiCl		
10mM	EDTA		
1%	LiDS		

DNA-

m
m 10 m
m 2.8 m 4
.5 m
5%

US-A-4336173

Qiagen Pharmacia

Serotec

Dyno Particles AS Lillestr m Norway

Sintef	EP-A-106873	D
YNABEADS	Dynal AS Oslo Norway	

4,336,173	4,459,378
-----------	-----------

4,654,267

DNA

RNA

DNA

DNA

10mM

-HCl pH 8.0/10mM NaCl

PCR

DNA

DNA

DNA

65

10

PCR

(14)

DNA

RNA	DNA
5' cap	5' cap
5' UTR	5' UTR
5' start codon	5' start codon
5' coding sequence	5' coding sequence
5' stop codon	5' stop codon
5' 3' UTR	5' 3' UTR
5' poly(A) tail	5' poly(A) tail

DNA

RNA

NaOH

RNA

DNA

RNA

RNA

DNA

DNA

RNA

DNA

RNA

RNA

RNA

RNA

RNA

DNA

RNA

DNA

RNA

DNA

RNA

D

NA

LiCl

RNA

GTC

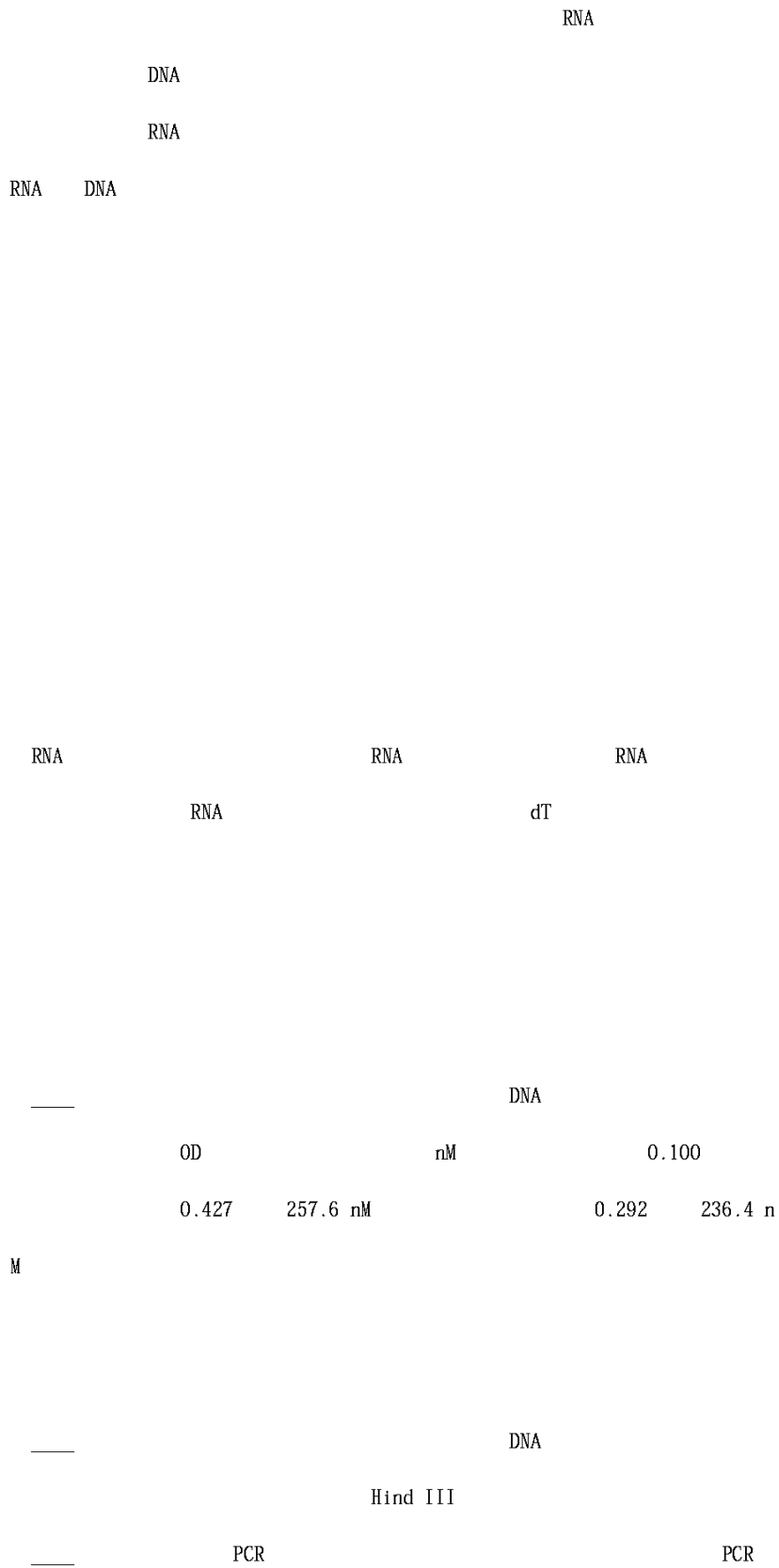
RNA

DNA

DNA

DNA

RNA



Hind III PCR

PCR Hin

d III

Hind III

DNA Dynabeads DNA DIRECT

DNA 10 1

DNA Dynabeads DNA DIRECT

Dynabeads DNA DIRECT

DNA

Hind III

II Dynabeads DNA DIRECT DNA

Dynabeads DNA DIRECT DNA 20 g

AMXY PCR

DNA 200 ng AMXY PCR

Dynabeads DNA DIRECT

Dynabeads DNA DIRECT 10 1

DNA 10% DNA

PCR 20% Hind

III M 100 bp L

EDTE

Dynabeads DNA DIRECT

A B 10 L

DNA

1 DNA 10% DNA

PCR 20%

20% DNA

—

EDTA Dynabeads DNA DIRECT

10 1 DNA

10% DNA PCR 20%

II

Dynabeads DNA DIRECT

10 1

DNA 10% DNA

PCR 20%

—

Dynabeads DNA DIRECT

1 Dynabeads

DNA DIRECT A B

DNA 10% DNA

PCR 20% II 10⁵

Daudi Dynabeads DNA DIRECT

DNA 120 1 DNA 1

PCR 20%

DNA Hind III PCR

100 bp

— 10 Dynabeads DNA DIRECT

A DNA

DIRECT DNA PCR 20%

M 100 bp B

20ng DNA C

PCR
 11 mRNA Dynabeads DNA DIRECT mRNA Dynabeads
 ads DNA DIRECT DNA Dynabeads Oligo(dT)₂₅
 100 DNA DIRECT Dynabeads
 DNA mg
 mg mg
 10mg mRNA
 DNA
 DNA
 RNA
 12 (A) (B) (C) (D)
 DNA PCR
 DNA 200 1 DNA DIRECT 20%
 DNA 10% RPC
 PCR 2.5% DNA
 5% 16S rRNA DNA
 DNA
 DNA 18S rDNA trn
 L B15C
 DNA
 PCR

 _____ DNA _____
 4 x 10⁶ HL 60 PBS
 10 1 PBS 0.1 ml 5% SDS/10 mM
 TrisCl pH 8.0/1 mM EDTA] に再懸濁させたトシル活性化 Dynabeads® M-
 280 DYNAL A/S
 ープすることにより得られる 1 mg の Dynabeads® M-280* を加えた。これに、

ml
とし、その後 DNA を結合した Dynabeads® を、磁石に引き付け、液相を除

ml 50mM NaCl/10mM TrisCl pH 8.0/
1mM EDTA DNA 0.1 ml

65

DNA

DNA OD₂₆₀ /OD₂₈₀ 1.72

TE DNA 1.7 1.9 DNA

OD₂₆₀

50 g/ml OD₂₆₀ =1.0 OD₂₆₀ 0.436 0
.1ml 10 mm 2.18 g DNA
DNA 2.67 g 82%
DNA >20 kb

表 1

PERKIN-ELMER LAMBDA BIO UV/VIS 分光器
アプリケーション番号 3 : 2 6 0 / 2 8 0 N M 比

試 料	サイクル	波 長	デ ー タ	単 位
	15:50	オートゼロ		
004	15:56	260.0 nm	0.436	ABS
		280.0 nm	0.253	ABS
		比	1.723	RAT

DNA

1 EDTA 50 1 5% SDS 1 PBS

の 50 μg の Dynabeads® M-280* を加えた。この溶解物を、1 分室温でイン

0.5 ml TrisCl pH 7.5

DNA

0.5 ml 10mM TrisCl pH 7.5

DNA 40 1 TE 10mM TrisCl pH 8.0/1 mM EDTA

1 PCR GAPDH PCR

PCR

50 1 PCR 10 1

+++ 溶解バッファ	DNA	洗浄バッファ	結 果
2% SDS		50 mM NaCl/1 x TE	+++
2% SDS/1 x TE		50 mM NaCl/1 x TE	+++
2% SDS/1 x TE/10 mM NaCl		50 mM NaCl/1 x TE	+++
5% SDS		50 mM NaCl/1 x TE	+++
5% SDS/1 x TE		50 mM NaCl/1 x TE	+++
5% SDS/1 x TE/10 mM NaCl		50 mM NaCl/1 x TE	+++
1% LiDS/10 x TE/0.5 M LiCl		50 mM NaCl/1 x TE	+++
1% LiDS/10 x TE/0.5 M LiCl		150 mM LiCl/1 x TE	+++
5% LiDS		150 mM LiCl/1 x TE	+++
5% SDS		150 mM LiCl/1 x TE	+++
1% サルコシル		150 mM LiCl/1 x TE	+++

1 x TE 10mM TrisCl pH 8.0/1 mM EDTA 10 X TE 100 mM TrisCl

pH 8.0/10 mM EDTA

実施例 1 の操作を辿ると、未被膜 Dynabeads® M-450 (Dyna1 A/S, オスロ、

CD2 DNA

50 1 50 1 PB

S 150mM NaCl/10mM Na₂HPO₄ pH 7.4 10 1 4 x 10⁶
 の Dynabeads® M-450 Pan-T (CD2)(Dynal AS, オスロ、ノルウェーより入手

30

/ 200 1 PBS 200
 g の Dynabeads® M-280* (同上) および 200 μ l の溶解バッファ [100mM

Tris-HCl pH 8.0/500mM LiCl/10mM EDTA pH 8.01/1% LiDS

DNA/

DNA/ 200 1 [10mM Tris-HCl

pH 8.0/150mM LiCl/1mM EDTA pH 8.0] 50 1

65

1 GAPDH PCR

 DNA

DNA ml EDTA Dynabeads DNA

DIRECT Dynal A/S,

Dynabeads® M-280* と等価のビーズを含有したキット)を用いて、同じ血液

10 1 DNA 65

DNA Dynabeads

Dynabeads DNA DIRECT DNA

DNA 0.2%

DNA 10 1 (5ml 0

.2%)

DNA John John S.W.M. G Weitzner R Rose

n C.R.scriver 1991 A Rapid Procedure for Extracting Genomic DNA
from Leukocytes Nucl Acid Res 19(2): 408

Dynabeads DNA DIRECT 200 1 Dynabeads DN
A DIRECT 10 1 1.5 ml
(/ 200 g Dynabeads)
DNA Dynabeads

DNA/Dynabeads Dynal E(Magnetic Particle
Collector E) (MPC-E)

Dynal MPC

10 1

TE pH 8.0

65

DNA

DNA

1.5%

1 x TAE

DS34

667

I

1

(

)

DNA

(

)

Hind III

23.13 kb

DNA

20 kb

DNA DIRECT

ACD

DNA

10%

DNA

200 ng

X-Y

(X-Y homologous amelogenin) (AMXY)

(Akane A. K. M

atsubara H Nakamura S Takahashi

K Kimura 1994 Purification

of Highly Degraded DNA by Gel Filtration for PCR BioTechniques 16(2):

235-238) amplicon PCR

DNA DIRECT DNA

PCR 50 1 10 x PCR Perkin Elmer

1 x dNTP Pharmacia 0.2mM

(amplitaq) (Perkin

Elmer pmol AMXY-1F 5'-CTGA

TGGTTGGCCTCAAGCCT-GTG-3' AMXY-4R 5'-TTCATTGTAAGAGCAAAGCAAACA-3'

PCR Perkin Elmer GeneAmp PCR System 9600

AMXY PCR 94 38 x[94 30

55 30 72] 72 10

50 1 PCR 10 1 1.5%

1 x TAE

DS34 667

II X-Y

DNA (Akane et al 1994

) 908 bp X 719 bp Y

II X Y

Dynabeads DNA DIRECT

DNA

DNA PCR DNA

DIRECT DNA

DNA PCR DNA DIRECT

DNA PCR 10

溶解／結合 バッファ:

0.5 M LiCl

1 % LiDS

0.1 M TrisCl pH 7.5

10 mM EDTA

5 mM ジチオトレイトール (DTT)

洗浄バッファ:

0.15 M LiCl

10 mM Tris-HCl pH 8.0

1 mM EDTA

DNA

Dynal AS

Dynabeads DNA DIRECT

DNA

A: 3.6×10^6 /ml B: 2.6×10^6 /ml

200 1 Dynabeads DNA DIRECT

1.5 ml

DNA Dynabeads

DNA/Dynabeads

Dynal

E(MPC-E)

Dynal MPC

40 1 TE pH 8.0

PCR

10%

DAPDH

amplicon

PCR

PCR

50

10 x PCR

(Perkin E

1mer) 1 x

dNTP Pharmacia 0.2mM

(amplitaq) (Perkin Elmer)

pmol GAPDH-Forward (5' -ACAGTCCATGCCATCAC
 TGCC-3') GAPDH-Reverse (5' -GCCTGCTTCACCACCTTCTTG-3')

PCR Perkin Elmer GeneAmp PCR System 9600 GAPDH

PCR 94 34 x[94 30 61 30

72] 72 10

DNA PCR 1.5%

50 1 10 1 D

NA 50% 1 x TAE

DS34 667

DNA

Dynabeads DNA DIRECT Dynal AS

EDTA

DNA

DNA 200 1 Dynabeads DNA DIRECT

1 10 1

1.5 ml

DNA Dynabeads

DNA/Dynabeads

Dynal

E (MPC-E)

Dynal MPC

20 40 1 TE pH 8.0 40

1 20 1

10%(20%)

(DAPDH) amplicon PCR

PCR Dynabeads TE

PCR 50 1 10 x PCR

Perkin Elmer 1 x dNTP

(Pharmacia) 0.2mM (

amplitaq)(Perkin Elmer) pmol G

APDH-Forward(5'-ACAGTCCATGCCATCACTGCC-3')

GAPDH-Reverse(5'-GCCTGCT

TCACCACCTTCTTG-3')

PCR Perkin Elmer GeneAmp PCR S

ystem 9600 GAPDH PCR 94

34 x[94 30 61 30 72] 72 10

50 1 10 1 DNA 25%

50% DNA PCR

1.5%

1 x TAE DS34

667

1 PCR

DNA 20% 10 1

10%

. M Georgesz A.M Lew 1993 FoLT PCR: A Simple PCR Protocol fo
 r Amplifying DNA Directly from Whole Blood BioTechniques 14(3): 238-243
)

DNA DIRECT ACD(11) CPD

DNA

Dynabeads DNA DIRECT Dynal AS

EDTA DNA

20 +4

DNA

200 1 Dynabeads DNA DIRECT

1.5 ml

DNA Dynabeads

DNA/Dynabeads Dynal E (MPC-E)

Dynal MPC

40 1 TE pH 8.0 PCR

Dynabeads TE

10%

DAPDH amplicon PCR

DNA PCR

1.5%				50	1		10	1	
DNA	50%								1 x TA
E			DS34						667
							I		
+4	-20								
						Dynabeads DNA DIRECT			
DNA									
10	1								
		1.5 ml		40	1	PBS			
		90		DNA		Dynabeads DNA DIRECT			
							10%		
	DAPDH		amplicon		PCR				
	DNA		PCR						1.5
%				50	1		10	1	
DNA	50%								1 x TAE
			DS34						667
							II		
			DNA		PCR				
<hr/>									
			DNA						
<hr/>									
	DNA								1
DNA DIRECT		DNA							
						200	1	Dynabeads DNA	
DIRECT							1.5ml		
	DNA		Dynabeads						
DNA/Dynabeads			Dyna1					E(MPC-E)	

Dyna1 MPC

40 1 TE pH 8.0 PCR

Dynabeads TE

10% DAPDH

amplicon PCR

DNA PCR 1.5%

50 1 10 1 D

NA 50% 1 x TAE

DS34 667

I

()

I DNA

1 DNA DIRECT

DNA

1

1 10 PCR

DNA

4 x 10⁵ Daudi cells DNA DIRECT

DNA 4 x 10⁵

DNA Dynabeads DNA DIRECT ml

ml

DNA/Dynabeads 120 1 TE

120 1 1 D

[illegible]

II 120

PCR

			DNA
Dynabeads	DNA DIRECT	Dyna1 AS	
			DNA

ds	200	1	Dynabeads	DNA DIRECT	1.5 ml
			DNA	Dynabeads	DNA Dynabea

DNA/Dynabeads Dynal E (MPC-E)

Dynal MPC

10

1	Dynabeads			
	DAPDH	amplicon		PCR
	PCR			1.5%
	50 1	10 1		

1 x TAE

DS34

667

10

PCR

mRNA

DNA DIRECT

DNA

mRNA

100

0.75ml

/

DNA DIRECT Dynabeads

DNA-Dynabeads

Dynal

MPC-E

10mg

DNA DIRECT

DNA

(11)

Dynal

mRNA DIRECT

mg

Dynabeads

(dT)₂₅

Dy

nabeads

mRNA

mRNA-Dynabead

MPC-E

Li

DS

0.75ml

mRNA-Dynabead

LiDS

mRNA

20 1

mM Tri

s-HCl pH 7.5

65

Dyn

abeads

1.0%

11

EtBr-

DNA

rRNA

RNA

11

10

DNA

mRNA

	DYNAL	mRNA
DNA	DNA	DNA DIRECT
DNA		
溶解／結合 バッファ:		0.5 M LiCl 1 % LiDS 0.1 M TrisCl pH 7.5 10 mM EDTA 5 mM ジチオトレイトール (DTT)
LiDS 含有洗浄バッファ:		0.15 M LiCl 0.1 % LiDS 10 mM Tris-HCl pH 8.0 1 mM EDTA
洗浄バッファ:		0.15 M LiCl 10 mM Tris-HCl pH 8.0 1 mM EDTA

DNA

PCR

DNA

(E.Coli)

Baceillus cereus

LB

37

(Agrobacterium

tumefaciens)

YEB

40

28

(Sambrook J et al

. 1989 Molecular Cloning: A Laboratory Manual 2nd ed. Cold Spring Ha

rbour Laboratory NY.)

(Prochlo

rthrix)

NIVA

14

18

20

Norwegian Institute of Water Research 1991

20

450,000

DNA

mg 20mg

DNA

Saccharomyces cerevisiae

IMR (Epp

ley R et al. 1967 Exp Mar Biol Ecol 1 191-208)

Arabidopsis thaliana

(Hordeum vulgare)

Perca fluviatilis

mg 30 100mg 100

400 mg DNA

DNA

forceps

(Kontes Scientific

Instruments Vineland New Jersey USA)

DNA

Dynabeads DNA DIRECT Dynal AS

DNA 200 1 D

ynabeads DNA DIRECT / 200 g Dynabeads

1.5 ml

15 DNA Dynabeads

65 15

DNA/Dynabeads Dynal E (MPC-E)

Dynal MPC

40 1 TE pH 8.0

65

DNA

DNA

1.5%

1 x TAE

DS34

667

溶解／結合バッファ:**0.5 M LiCl****1% LiDS****0.1 M TrisCl pH 7.5****10 mM EDTA****5 mM ジチオトレイトール (DTT)****洗浄バッファ:****0.15 M LiCl****10 mM Tris-HCl pH 8.0****1 mM EDTA**

DNA DIRECT

/

DNA

Sambrook J et al. 1989

A-5 Sigma Chemicals Co.

St Louis USA

DNA Scot O.R Bendich A.J. 1994 "Plant Molecular Biol
ogy Manual" page D1: 1-8 Kluwer Academic Publisher Belgium

PCR

DNA

DNA

PCR

DNA

PC

R

PCR

15 pmol

200 M

dNTP 10mM

Tris-HCl pH 8.8 1

.5 mM

MgCl₂ 50 mM

KCl 0.1% Triton X-100

DynaZyme

Finnzymes Oy Finland

0.1-5 1

DN

A

50 1

PCR

Perkin Elmer GeneAmp PCR Sy

stem 9600

amplicon			
PCR	94-97	DNA	72

amplicon	IUD
Brosius J. et al. 1978 Proc Natl Acad Sci. USA 57 4801	
-4805	E Coli 334 939 16S rRNA

プライマー: CC 5'-TGTAACGACGGCCAGTCCAGACTCCTACGGGAGGCAGC-3'
CD 5'-CTTGTGCGGGCCCCCGTCAATTC-3'

CC	-21 M13	5
DNA		: 96 1
5	70	30

18S rRNA	Medlin et al. 1990 Gene 71 491-499
A	B
94	30 50 72 35

18S rRNA	600 bp.	White et al. 1990 Innis M.A et al.
"PCR Products a Guide to Methods and Applications" page 315-32		
2 Academic Press New York	NS3	NS4

94	30	53	30	72
94	30	50	30	15
72		25		

	tRNA	I		Fangan et al. 1994	BioTechn
iques 16	484-494		C	D	
94	30	55	30	72	30

Arabidopsis thaliana B15C 800 bp

5'-CGGGATCCCTAGGAGACACGGTGCCG-3' および
5'-GGAATTCGATCGGCGGTCTTGAAAC-3'

94	30	59	30	72	35
----	----	----	----	----	----

Barley B15C 800 bp

5'-CGGATCCCGTCATCCTTCTCTCGCACCCC-3' および
5'-GGAATTCCTTCTTGGAGGGCAGGTCGGCG-3'.

94	30	60	30	72	35
----	----	----	----	----	----

_____(Perch)

	D-	800-900 bp	Hoelzel et al. 1
991	Mol Biol Evol.	8 475-493	HV2

5'-GGTGACTTGCATGTGTAAGTTCA-3'.

96	52	72	30
----	----	----	----

1.5%

1 x TAE DS34

667

<hr/>		100-1000 ng	
DNA	(12A)		
		65	15
DNA	500 ng	mg	
0.25%	DNA		
<hr/>			
DNA	100-200 ng		
300-500ng	(12B)		
DNA			5
%	DNA	PCR	(12B)
		0.5-5%	DNA PCR
<hr/>			
		DNA DIRECT	
		200-400 ng	DNA (12C)
PCR	DNA	DNA	PCR
5%	DNA		
<hr/>			
		DNA	
		65	15
(12D)	PCR	DNA	DNA
		PCR	5% DNA
<hr/>			
300-500 ng	DNA		
		()	DNA 5
%	DNA		

PCR

Hultman et

al. 1989 Nucleic Acids Res. 17 4937-4946

⁶ 実施例 3 に記載したアンプリコン

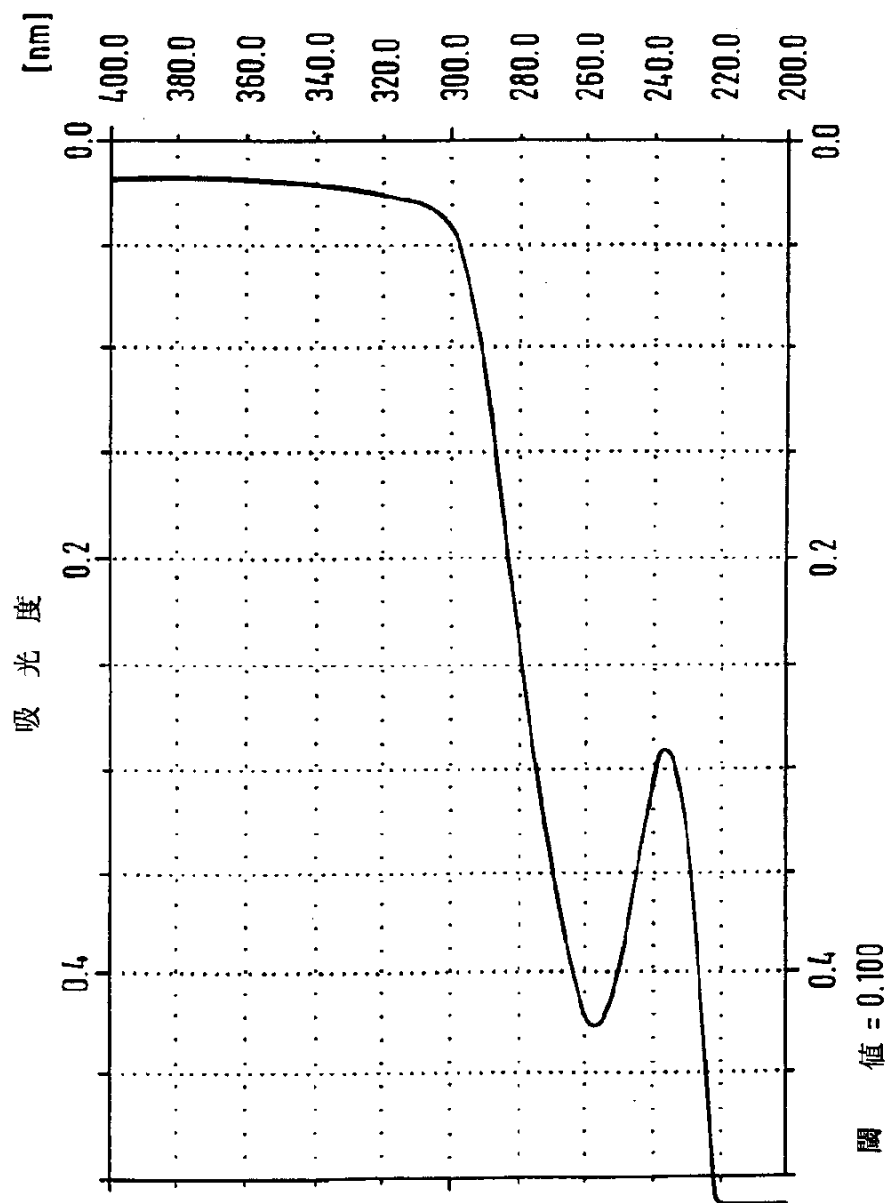
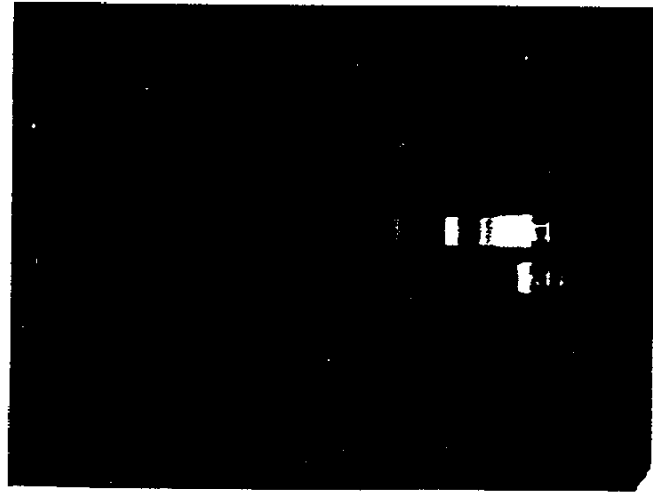


FIG. 1



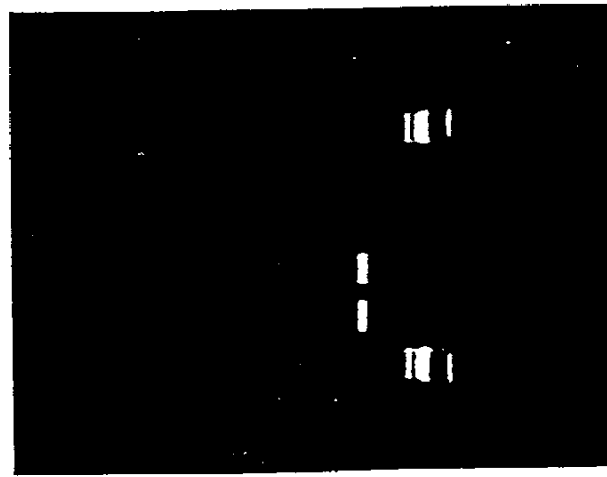
1 2

FIG. 2



1 2 3

FIG. 3



1 2 3 4 5 6

FIG. 4

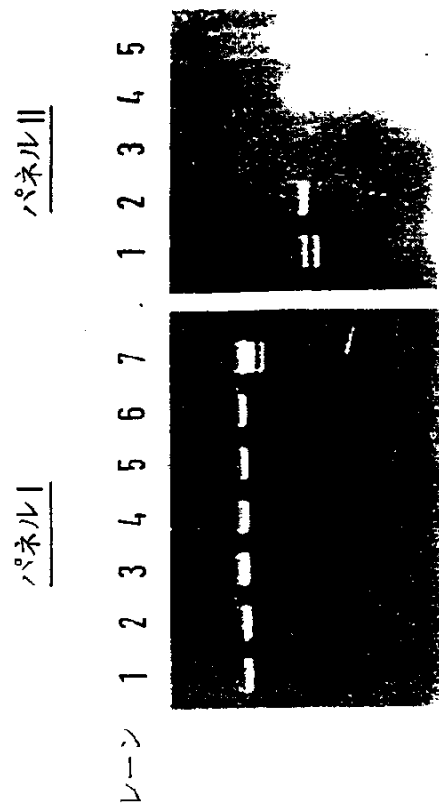


FIG. 5

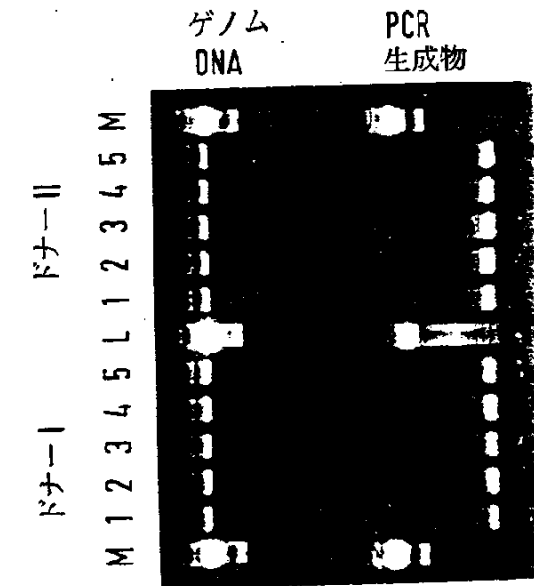


FIG. 6

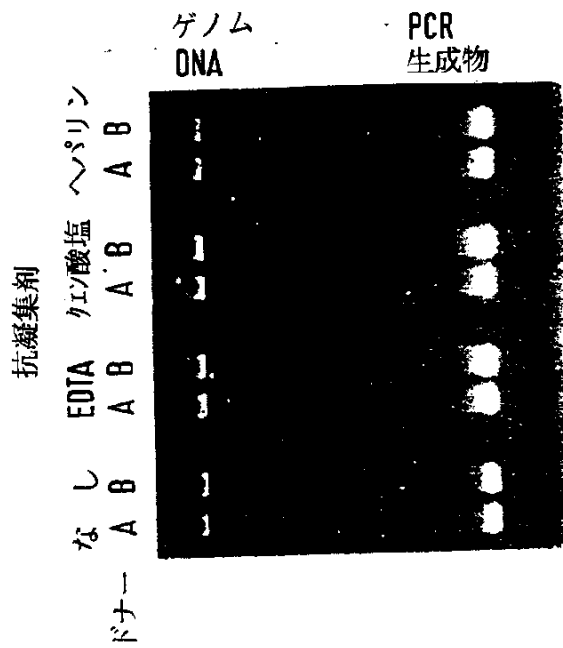


FIG. 7

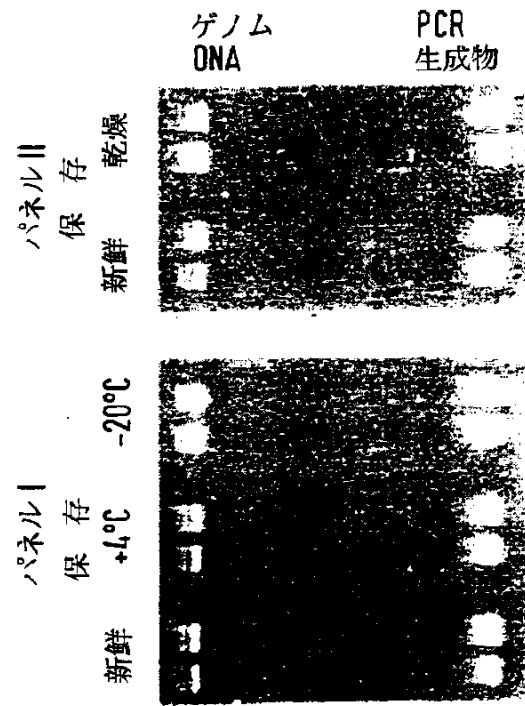


FIG. 8

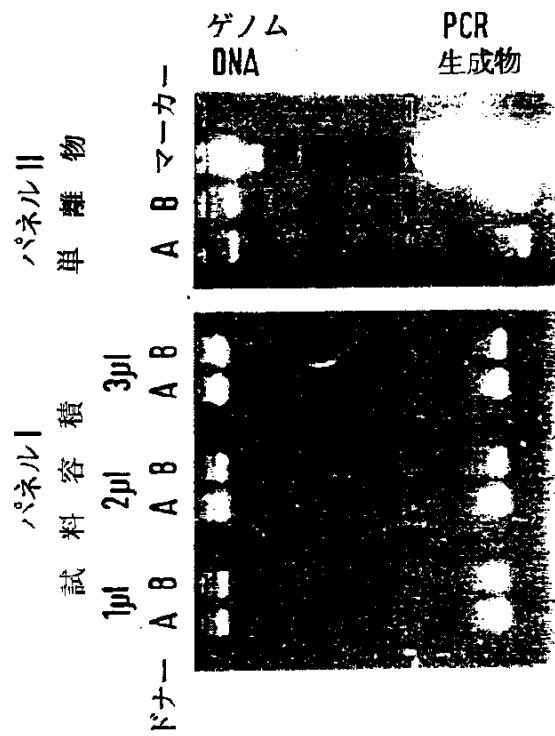


FIG. 9

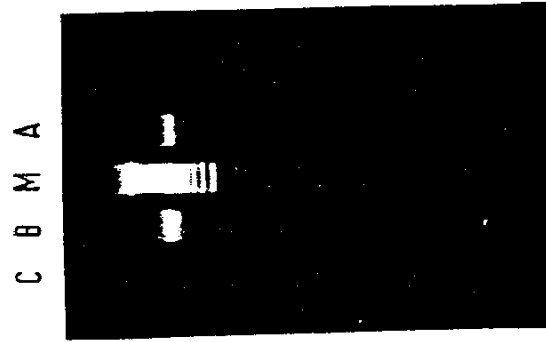


FIG. 10

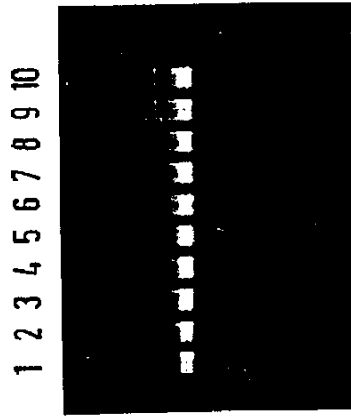


FIG. 11

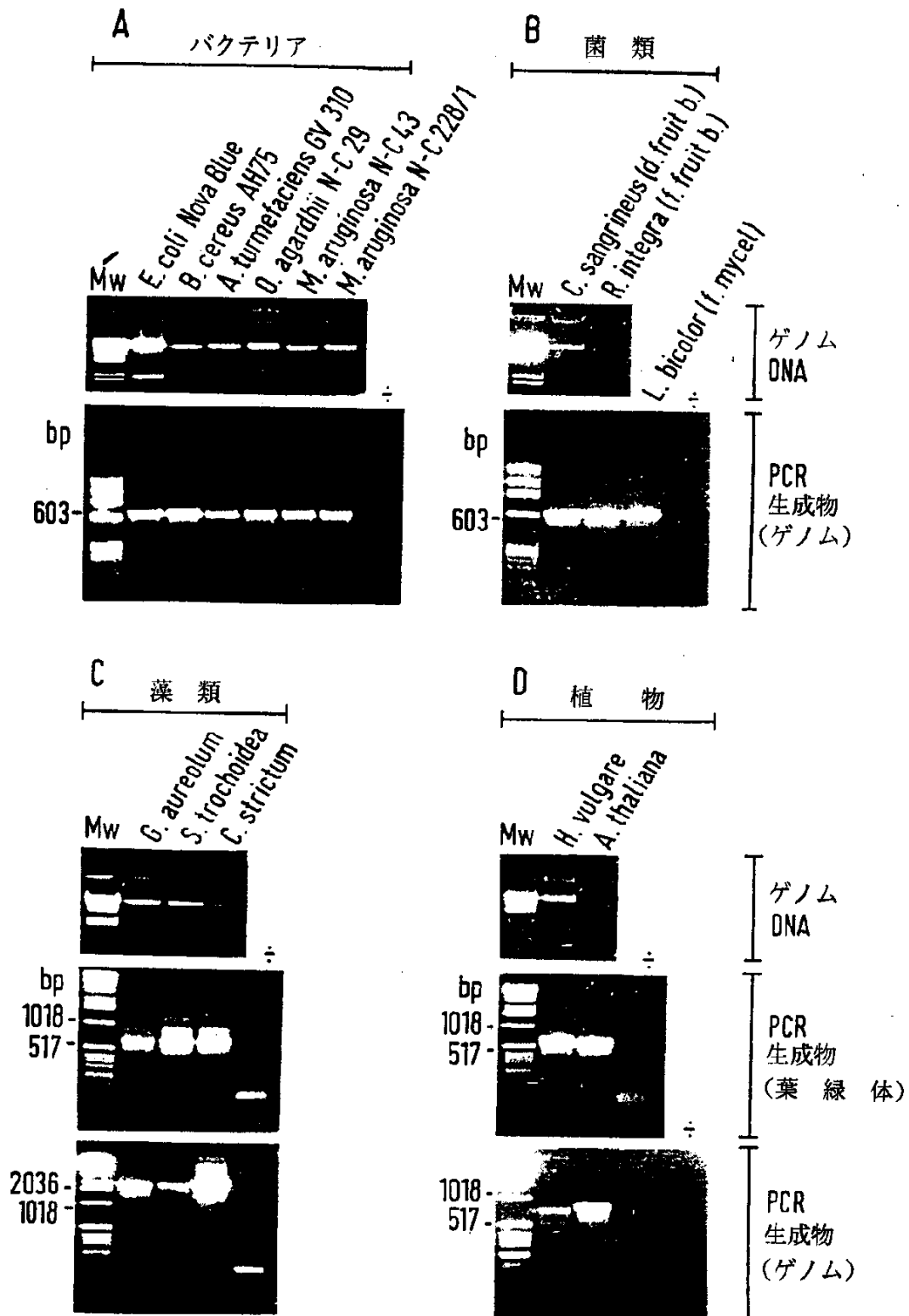


FIG. 12

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/GB 95/02893

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/10

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO, A, 93 25912 (MEDICAL RESEARCH COUNCIL) 23 December 1993 *see the whole patent* ---	1-18
X	JOURNAL OF APPLIED BACTERIOLOGY, vol. 74, 1993, pages 78-85, XP002007385 K. SMALLA ET AL.: "Rapid DNA extraction protocol from soil for polymerase chain reaction mediated amplification" *see the whole article* --- -/--	1-18

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

* Special categories of cited documents:

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- *E* earlier document but published on or after the international filing date
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- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
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- *Z* document member of the same patent family

Date of the actual completion of the international search

3 July 1996

Date of mailing of the international search report

02.08.96

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INTERNATIONAL SEARCH REPORT

International Application No.

PCT/GB 95/02893

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>APPLIED AND ENVIRONMENTAL MICROBIOLOGY, vol. 60, no. 5, 1994, pages 1572-1580, XP002007386 N.I. MORE ET AL.: "Quantitative cell lysis of indigenous microorganisms and rapid extraction of microbial DNA from sediment" *see the whole article* ---</p>	1-18
X	<p>EMBO JOURNAL, vol. 4, no. 4, 1985, pages 913-918, XP002007387 D.A. JACKSON ET AL.: "A general method for preparing chromatin containing intact DNA" *see the whole article* ---</p>	1-18
X	<p>EMBO JOURNAL, vol. 3, no. 8, 1984, pages 1837-1842, XP002007388 P.R. COOK : "A general method for preparing intact nuclear DNA" *see the whole article* ---</p>	1-18
X	<p>EXPERIMENTAL CELL RESEARCH, vol. 190, 1990, pages 294-296, XP002007389 G. KONAT ET AL.: "Rapid isolation of genomic DNA from animal Tissues" *see the whole article* ---</p>	1-18
X	<p>ANALYTICAL BIOCHEMISTRY, vol. 164, 1987, pages 207-213, XP002007390 P.M. GLEE ET AL.: "Methods for DNA extraction from Candida albicans" *see the whole article* -----</p>	1-8

INTERNATIONAL SEARCH REPORT

Information on patent family members

Int. l. Application No

PCT/GB 95/02893

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
W0-A-9325912	23-12-93	AU-B- 4343993	04-01-94
		AU-B- 4344093	04-01-94
		W0-A- 9325709	23-12-93

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